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(54) Alkaline cellulase variants

(57) Provided in the present invention are an alkaline cellulase variant obtained by using a cellulase having an amino acid sequence exhibiting at least 90% homology with the amino acid sequence represented by SEQ. ID NO:1 and substituting the amino acid residue of the cellulase at (a) position 10, (b) position 16, (c) position 22, (d) position 33, (e) position 39, (f) position 76, (g) position 109, (h) position 242, (i) position 263, (j) position 308, (k) position 462, (l) position 466, (m) position 468, (n) position 552, (o) position 564, or (p) position

608 in SEQ. ID NO:1, or at a position corresponding thereto with another amino acid residue; a gene encoding the variant; a vector containing the gene; a transformant containing the vector; and a detergent composition containing the alkaline cellulase variant.

The present invention makes it possible to provide an alkaline cellulase which acts favorably in an alkaline range, and can be mass produced easily because it has a high secretion capacity or enhanced specific activity.

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Description**Technical Field**

- 5 [0001] The present invention relates to alkaline cellulase variants which can be incorporated in laundry detergents and the like.

Background Art

- 10 [0002] Cellulases are enzymes which were considered to act only in a neutral or acid range but not in an alkaline laundry detergent solution. Finding of an alkaline cellulase derived from alkaliphilic microorganisms belonging to *Bacillus* sp. by Horikoshi (refer to Japanese Patent Publication No. Sho 50-28515 and Horikoshi & Akiba, Alkaliphilic Microorganisms, Springer, Berlin, 1982) enabled its use for laundry heavy-duty detergents. Since then, alkaline cellulases produced by alkaliphilic microorganisms belonging to *Bacillus* sp. have been developed (refer to Japanese Patent Publication No. Sho 60-23158, Japanese Patent Publication No. Hei 6-030578 and U.S. Patent No. 4945053) and are now incorporated in laundry detergents.

[0003] Recent progress in genetic engineering has enabled mass production of enzymes for detergents, which also applies to the production of alkaline cellulases. A number of genes for alkaline cellulases have already been cloned, their nucleotide sequences have been determined.

- 20 Furthermore, technique for mutagenesis and breeding of the enzyme-producing bacteria or mutagenesis of a gene encoding the enzyme has been introduced.

[0004] Productivity of them on an industrial scale however does not reach a satisfactory level, and there is a demand for alkaline cellulases which can be produced efficiently.

- 25 [0005] An object of the present invention is therefore to provide an alkaline cellulase which acts favorably in an alkaline region, and can be mass produced readily because of having high secretion capacity or having enhanced specific activity.

Disclosure of the Invention

- 30 [0006] The present inventors have searched a novel alkaline cellulase which can be produced efficiently without losing its inherent properties. As a result, it has been found that an alkaline cellulase variant, wherein an amino acid residue at a specific position of an amino acid sequence having at least 90% homology with the amino acid sequence represented by SEQ. ID NO:1 has been substituted with another amino acid residue, can be mass produced because it acquires high secretion capacity or enhanced specific activity.

- 35 [0007] In one aspect of the present invention, there are thus provided an alkaline cellulase variant obtained by substituting the amino acid residue of a cellulase having an amino acid sequence exhibiting at least 90% homology with the amino acid sequence represented by SEQ. ID NO:1 at (a) position 10, (b) position 16, (c) position 22, (d) position 33, (e) position 39, (f) position 76, (g) position 109, (h) position 242, (i) position 263, (j) position 308, (k) position 462, (l) position 466, (m) position 468, (n) position 552, (o) position 564, or (p) position 608 in SEQ. ID NO:1 or at a position corresponding thereto with another amino acid residue; and a gene encoding the alkaline cellulase variant.

- 40 [0008] In another aspect of the present invention, there are also provided a vector containing the gene and a transformant containing the vector.

[0009] In a further aspect of the present invention, there is also provided a detergent composition having the above-described alkaline cellulase variant incorporated therein.

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Brief Description of the Drawing**[0010]**

- 50 FIG. 1 shows aligned amino acid sequences of cellulases having at least 90% homology with the amino acid sequence represented by SEQ. ID NO: 1.

Best Mode for Carrying Out the Invention

- 55 [0011] The alkaline cellulase variants according to the present invention are obtained by using, as a cellulase to be mutated (which may hereinafter be called "parent alkaline cellulase"), a cellulase having an amino acid sequence exhibiting at least 90% homology with the amino acid sequence represented by SEQ. ID NO:1 and replacing the amino acid residue of the parent alkaline cellulase at the position (a) to (p) in the above-described SEQ. ID. NO:1 or at a

position corresponding thereto with another amino acid residue. The alkaline cellulase variants may be wild-type variants or artificial variants.

[0012] Alkaline cellulases having an amino acid sequence exhibiting at least 90% homology with the amino acid sequence represented by SEQ. ID NO:1 and serving as a parent alkaline cellulase embrace alkaline cellulases having the amino acid sequence represented by SEQ. ID NO: 1 and alkaline cellulases having an amino acid sequence exhibiting at least 90% homology with the above-described amino acid sequence. They may be either wild type alkaline cellulases or wild type variants. It is preferred that they have a molecular weight, as determined by the sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) or gel filtration, of $86,000 \pm 2,000$; have the optimal pH value of 7.5 to 9.5 when carboxymethyl cellulose is used as a substrate; have the optimal temperature of 40 to 50°C; favorably degrade lichenan in addition to carboxymethyl cellulose; and are sufficiently stable to incubation at pH 9 at 50°C for 10 minutes. Particularly preferred are cellulases having a molecular weight of $86,000 \pm 2,000$ (as determined by SDS-PAGE or gel filtration through a Sephacryl S200 column); having the optimum reaction pH of from 8.6 to 9.0; having the optimum reaction temperature of 50°C; capable of favorably degrading lichenan as well as carboxymethyl cellulose; and being recognized to have residual activity of 95% or greater (assuming that the residual activity after treatment at 30°C for 10 minutes is 100%) after treatment at 50°C for 10 minutes at pH 9 in the presence of 5 mM calcium chloride.

[0013] Examples of the "alkaline cellulase having the amino acid sequence represented by SEQ. ID NO:1" include Eg1-237 [derived from *Bacillus* sp. strain KSM-S237 (FERM BP-7875), Hakamada, et al., Biosci. Biotechnol. Biochem., 64, 2281-2289, 2000]. Examples of the "alkaline cellulase having an amino acid sequence exhibiting at least 90% homology with the amino acid sequence represented by SEQ. ID NO:1" include alkaline cellulases having an amino acid sequence exhibiting preferably at least 95% homology, more preferably at least 98% homology, with the amino acid sequence represented by SEQ. ID NO:1. Specific examples include alkaline cellulase derived from *Bacillus* sp. strain 1139 (Eg1-1139) (Fukumori, et al., J. Gen. Microbiol., 132, 2329-2335) (91.4% homology), alkaline cellulases derived from *Bacillus* sp. strain KSM-64 (Eg1-64) (Sumitomo, et al., Biosci. Biotechnol. Biochem., 56, 872-877, 1992) (homology: 91.9%), and cellulase derived from *Bacillus* sp. strain KSM-N131 (Eg1-N131b) (Japanese Patent Application No. 2000-47237) (homology: 95.0%).

[0014] The homology of an amino acid sequence can be calculated using a program such as maximum matching or search homology of GENETYX-WIN (Software Development Co.).

[0015] The amino acid residues at positions (a) to (p) of the parent alkaline cellulase having the amino acid sequence represented by SEQ. ID NO:1 are (a) leucine at position 10, (b) isoleucine at position 16, (c) serine at position 22, (d) asparagine at position 33, (e) phenylalanine at position 39, (f) isoleucine at position 76, (g) methionine at position 109, (h) glutamine at position 242, (i) phenylalanine at position 263, (j) threonine at position 308, (k) asparagine at position 462, (l) lysine at position 466, (m) valine at position 468, (n) isoleucine at position 552, (o) isoleucine at position 564 and (p) serine at position 608.

[0016] In the parent alkaline cellulases having an amino acid sequence exhibiting at least 90% homology with the amino acid sequence represented by SEQ. ID NO:1, the amino acid residues at the positions corresponding to the above-described (a) to (p) of SEQ. ID NO: 1 are preferably (a) leucine at position 10, (e) phenylalanine at position 39, (f) isoleucine at position 76, (h) glutamine at position 242, (i) phenylalanine at position 263, (k) asparagine at position 462, (l) lysine at position 466, (m) valine at position 468 and (n) isoleucine at position 552, of which the parent alkaline cellulase having, in addition to the above-described amino acid residues, (b) isoleucine at position 16 and (c) serine at position 22 are more preferred and that having, in addition to these residues, (d) asparagine at position 33, (g) methionine at position 109, (j) threonine at position 308, (o) isoleucine at position 564 and (p) serine at position 608 are still more preferred.

[0017] Accordingly, of the parent alkaline cellulases having an amino acid sequence exhibiting at least 90% homology with the amino acid sequence represented by SEQ. ID NO:1, preferred are cellulases having the above-described enzymatic properties and/or having an amino acid sequence exhibiting preferably at least 95%, more preferably at least 98% homology with the amino acid sequence represented by SEQ. ID NO: 1, and moreover having, as amino acid residues at positions corresponding to the positions (a) to (p) of SEQ. ID NO:1, the above-described ones. Particularly preferred are cellulases having the above-described enzymatic properties, having an amino acid sequence exhibiting preferably at least 95%, more preferably at least 98% homology with the amino acid sequence represented by SEQ. ID NO: 1, and moreover having, as amino acid residues at positions corresponding to the positions (a) to (p) of SEQ. ID NO:1, the above-described ones.

[0018] When a cellulase having the amino acid sequence represented by SEQ. ID NO:1 is used as a parent alkaline cellulase, the alkaline cellulase variant of the present invention has an amino acid residue which is substituted for the amino acid residue at any one of the positions (a) to (p), while a cellulase having an amino acid sequence exhibiting at least 90% homology with the amino acid sequence represented by SEQ. ID NO: 1 (except the alkaline cellulase represented by SEQ ID NO:1) is used as a parent alkaline cellulase, the amino acid residue at a position corresponding to any one of the above-described positions (a) to (p) of SEQ. ID NO: 1 has been substituted with another amino acid.

[0019] As such another amino acid residue, glutamine, alanine, proline or methionine, especially glutamine is preferred at position (a), asparagine or arginine, especially asparagine is preferred at position (b), proline is preferred at position (c), histidine is preferred at position (d), alanine, threonine or tyrosine, especially alanine is preferred at position (e), histidine, methionine, valine, threonine or alanine, especially histidine is preferred at position (f), isoleucine, leucine, serine or valine, especially isoleucine is preferred at position (g), alanine, phenylalanine, valine, serine, aspartic acid, glutamic acid, leucine, isoleucine, tyrosine, threonine, methionine or glycine, especially alanine, phenylalanine or serine is preferred at position (h), isoleucine, leucine, proline or valine, especially isoleucine is preferred at position (i), alanine, serine, glycine or valine, especially alanine is preferred at position (j), threonine, leucine, phenylalanine or arginine, especially threonine is preferred at position (k), leucine, alanine or serine, especially leucine is preferred at position (l), alanine, aspartic acid, glycine or lysine, especially alanine is preferred at position (m), methionine is preferred at position (n), valine, threonine or leucine, especially valine is preferred at position (o) and isoleucine or arginine, especially isoleucine is preferred at position (p).

[0020] The "amino acid residue at a position corresponding thereto" can be identified by comparing amino acid sequences by using known algorithm, for example, that of Lipman-Pearson's method, and giving a maximum similarity score to the multiple regions of similarity in the amino acid sequence of each alkaline cellulase. The position of the homologous amino acid residue in the sequence of each cellulase can be determined, irrespective of insertion or deletion existing in the amino acid sequence, by aligning the amino acid sequence of the cellulase in such manner (Fig. 1). It is presumed that the homologous position exists at the three-dimensionally same position and it brings about similar effects with regard to a specific function of the target cellulase.

[0021] With regard to another alkaline cellulase having an amino acid sequence exhibiting at least 90% homology with SEQ. ID NO:1, specific examples of the positions corresponding to (a) position 10, (b), position 16, (c) position 22, (d) position 33, (e) position 39, (f) position 76, (g) position 109, (h) position 242, (i) position 263, (j) position 308, (k) position 462, (l) position 466, (m) position 468, (n) position 552, (o) position 564 and (p) position 608 of the alkaline cellulase (Eg1-237) represented by SEQ. ID NO: 1 and amino acid residues at these positions will be shown below.

Table 1

	Egl-237	Egl-1139	Egl-64	Egl-N131b
(a)	10Leu	10Leu	10Leu	10Leu
(b)	16Ile	16Ile	16Ile	Nothing corresponding thereto
(c)	22Ser	22Ser	22Ser	Nothing corresponding thereto
(d)	33Asn	33Asn	33Asn	19Asn
(e)	39Phe	39Phe	39Phe	25Phe
(f)	76Ile	76Ile	76Ile	62Ile
(g)	109Met	109Met	109Met	95Met
(h)	242Gln	242Gln	242Gln	228Gln
(i)	263Phe	263Phe	263Phe	249Phe
(j)	308Thr	308Thr	308Thr	294Thr
(k)	462Asn	461Asn	461Asn	448Asn
(l)	466Lys	465Lys	465Lys	452Lys
(m)	468Val	467Val	467Val	454Val
(n)	552Ile	550Ile	550Ile	538Ile
(o)	564Ile	562Ile	562Ile	550Ile
(p)	608Ser	606Ser	606Ser	594Ser

[0022] These amino acid residues may be substituted at two or more positions simultaneously as long as the substitution does not bring about any change in enzyme activity or enzymatic characteristics. Below-described are preferred specific examples of the case where substitution is carried out at two or more positions simultaneously. The amino acid is expressed by three letters, and "+" means that substitution at one position is followed by another substitution, while "/" means that any amino acid indicated thereby is usable.

[0023] Preferred double substitution examples include Leu10(Gln/Ala/Pro/Met) + Ile16(Asn/Arg), Ile16(Asn/Arg) +

Ser22Pro, Leu10(Gln/Ala/Pro/Met) + Ser22Pro, Asn33His + Phe39(Thr/Tyr/Ala), Leu10(Gln/Ala/Pro/Met) + Gln242(Ser/Ala/Phe/Val/Asp/Glu/Gly), Ile16(Arg/Asn) + Ser22Pro, Ser22Pro + Gln242(Ala/Ser/Phe/Val/Ile/Gly/Glu/Asp/Thr/Leu/Met/Tyr), Gln242(Ala/Ser/Phe/Val/Ile/Gly/Glu/Asp/Thr/Leu/Met/Tyr) + Phe263(Ile/Leu/Pro/Val), Leu10(Gln/Ala/Pro/Met) + Thr308(Ala/Ser/Gly/Val), Ile16(Asn/Arg) + Asn462(Thr/Leu/Phe/Arg), Ser22Pro + Val468(Ala/Asp/Gly/Lys),
 5 Asn33His + Ile552Met, Asn33His + Ile564(Val/Thr/Leu), and Gln242(Ala/Ser/Phe/Val/Ile/Gly/Glu/Asp/Thr/Leu/Met/Tyr) + Ser608(Ile/Arg), with Leu10Gln + Ser22Pro, Asn33His + Phe39Ala, Ser22Pro + Gln242Ala, Ser22Pro + Gln242Phe, and Ser22Pro + Gln242Ser being particularly preferred.

[0024] Preferred triple substitution examples include Leu10(Gln/Ala/Pro/Met) + Ser22Pro + Gln242(Ala/Ser/Phe/Val/Ile/Gly/Glu/Asp/Thr/Leu/Met/Tyr), Ile16(Asn/Arg) + Ser22Pro + Gln242(Ala/Ser/Phe/Val/Ile/Gly/Glu/Asp/Thr/Leu/Met/Tyr), and Ile76(His/Met/Val/Thr/Ala) + Gln242(Ala/Ser/Phe/Val/Ile/Gly/Glu/Asp/Thr/Leu/Met/Tyr) + Lys466(Leu/Ala/Ser), with Leu10Gln + Ser22Pro + Gln242Ala, Ile16Asn + Ser22Pro + Gln242Ser, and Ile16Asn + Ser22Pro + Gln242Phe being especially preferred.

[0025] Preferred quadruple substitution examples include Leu10(Gln/Ala/Pro/Met) + Ser22Pro + Ile76(His/Met/Val/Thr/Ala) + Lys466(Leu/Ala/Ser), Leu10(Gln/Ala/Pro/Met) + Ile16(Asn/Arg) + Ile76(His/Met/Val/Thr/Ala) + Lys466(Leu/Ala/Ser), and Ile16(Asn/Arg) + Met109(Ile/Leu/Ser/Val) + Gln242(Ala/Ser/Phe/Val/Ile/Gly/Glu/Asp/Thr/Leu/Met/Tyr) + Ile564(Val/Thr/Leu), with Leu10Gln + Ser22Pro + Ile76His + Lys466Leu and Leu10Gln + Ser22Pro + Gln242Ala + Lys466Leu, and Leu10Gln + Ser22Pro + Gln242Ser + Lys466Leu being especially preferred.

[0026] Preferred quintuple substitution examples include Leu10(Gln/Ala/Pro/Met) + Ile16(Asn/Arg) + Ser22Pro + Ile76(His/Met/Val/Thr/Ala) + Lys466(Leu/Ala/Ser), and Ile16(Asn/Arg) + Gln242(Ala/Ser/Phe/Val/Ile/Gly/Glu/Asp/Thr/Leu/Met/Tyr) + Thr308(Ala/Ser/Gly/Val) + Ile552Met + Ser608(Ile/Arg), with Leu10Gln + Ile16Asn + Ile76His + Gln242Ser + Lys466Leu and Leu10Gln + Ile16Asn + Ile76His + Gln242Ala + Lys466Leu being especially preferred.

[0027] The alkaline protease variants of the present invention may be substituted simultaneously at positions greater in number than the above, for example, at 6 to 16 positions.

[0028] The alkaline cellulase variants of the present invention embrace, in addition to those obtained by replacing the amino acid residue at a position corresponding to any one of the above-described positions (a) to (p) with another amino acid, those having one to several amino acid residues depleted, replaced or added at the other position(s) of the amino acid sequence so far as they do not lose their alkaline cellulase activity.

[0029] The alkaline cellulase variants of the present invention are obtainable, for example, by the following method.

[0030] Specifically, it can be obtained by conducting substitution (which may be called "mutation") to a gene (SEQ. ID NO: 2) encoding a cloned parent alkaline cellulase (for example, an alkaline cellulase having the amino acid sequence represented by SEQ. ID No:1), transforming a proper host by using the resulting mutant gene, culturing the resulting recombinant host, and then collecting the target enzyme from the culture broth.

[0031] Cloning of a gene encoding the parent alkaline cellulase may be carried out using ordinarily employed recombinant DNA technology, for example, by the shotgun method or PCR method from a chromosomal DNA of the *Bacillus* sp. strain KSM-S237.

[0032] For mutagenesis of the gene encoding the parent alkaline cellulase, either ordinarily employed random mutagenesis or site-specific mutagenesis can be adopted. More specifically, mutagenesis can be effected using, for example, "Site-Directed Mutagenesis System Mutan-Super Express Km kit" (product of Takara Bio). An arbitrary sequence of the gene can be replaced with a sequence of another gene corresponding to the arbitrary sequence by using recombinant PCR (polymerase chain reaction) method (PCR protocols, Academic Press, New York, 1990).

[0033] The production of the alkaline cellulase variant of the present invention using the resulting mutant gene can be carried out by introducing the mutant gene in a DNA vector which permits stable expression of the enzyme, and then, transforming the host bacteria using the resulting recombinant vector.

[0034] When *Escherichia coli* is used as a host, examples of such a vector include pUC18, pBR322, and PHY300PLK (product of Yakult Honsha), and when *Bacillus subtilis* is used as a host, examples of the vector include pUB110, pHSP64 (Sumitomo, et al., Biosci. Biotechnol. Biochem., 59, 2172-2175, 1995) and PHY300PLK.

[0035] For the transformation of the host bacteria, protoplast method, competent cell method or electroporation method may be used. As the host bacteria, gram positive bacteria such as those belonging to *Bacillus* sp. (*Bacillus subtilis*), gram negative bacteria such as *Escherichia coli*, *Actinomyces* such as *Streptomyces*, yeasts such as *Saccharomyces* and fungi such as *Aspergillus*.

[0036] The transformant thus obtained may be cultured under proper conditions by using a culture broth containing an assimilable carbon source, nitrogen source, metal salt and vitamin. The enzyme is separated from the culture broth and then purified by an ordinarily employed method, followed by lyophilization, spray drying and/or crystallization to give the enzyme in a desired form.

[0037] The alkaline cellulase variant of the present invention thus obtained is improved in secretion capacity of the enzyme or is improved in the specific activity of the enzyme without losing the properties of the parent alkaline cellulase.

[0038] The term "improved in secretion capacity" as used herein means that when the parent alkaline cellulase and the alkaline cellulase variant are produced under similar conditions (for example, shake culture at 30°C for 72 hours

in a medium (PSM medium) composed of 3% (w/v) "Polypepton S" (product of Nihon Pharmaceutical), 0.5% fish meat extract (product of Wako Pure Chemicals), 0.05% yeast extract, 0.1% monopotassium phosphate, 0.02% magnesium sulfate 7 hydrate, tetracycline (15 µg/mL) and 5% maltose), and the enzyme activity and protein content in their culture supernatants are measured, the alkaline cellulase variant shows at least a predetermined improvement in the enzyme activity or protein content compared with those of the parent alkaline cellulase. For example, it means that at least 5%, desirably at least 10%, more desirably at least 20% increase in the enzyme activity or protein content recognized.

[0039] When any change in the specific activity is not recognized, any one of the enzyme activity and protein content may be measured, because the parent alkaline cellulase and alkaline cellulase variant are considered to have the same ratio of the enzyme activity to the protein content.

[0040] Accordingly, the alkaline cellulase variants of the present invention are useful as an enzyme to be incorporated in various detergent compositions.

[0041] Although there is no particular limitation on the amount of the alkaline cellulase variant of the present invention to be added to a detergent composition insofar as the cellulase can exhibit its activity, it is added preferably in an amount of from 0.0001 to 5 wt.%, more preferably from 0.00005 to 2.5 wt.%, still more preferably from 0.01 to 2 wt.% based on the detergent composition. When the alkaline cellulase variant is used in the form of granule, the content of the enzyme in the granule is preferably 0.01 to 50 wt.%, more preferably from 0.05 to 25 wt.%, still more preferably from 0.1 to 20 wt.%.

[0042] The detergent composition of the present invention preferably contains, in addition to the above-described alkaline cellulase (granules), a surfactant and a builder. As the surfactant, an anionic surfactant, nonionic surfactant, amphoteric surfactant and cationic surfactant may be used either singly or in combination, but an anionic surfactant and nonionic surfactant are preferred.

[0043] Preferred examples of the anionic surfactant include sulfate salts of a C₁₀₋₁₈ alcohol, sulfate salts of an alkoxylated C₈₋₂₀ alcohol, alkylbenzene sulfonate salts, alkylsulfate salts, paraffin sulfonate salts, α-olefin sulfonate salts, α-sulfo fatty acid salts, alkyl ester salts of an α-sulfo fatty acid, and fatty acid salts. In the present invention, linear alkylbenzene sulfonate salts having a C₁₀₋₁₄, more preferably C₁₂₋₁₄ alkyl straight chain are particularly preferred. As the counter ion of such a salt, alkali metal salts and amines are preferred, of which sodium and/or potassium, monoethanolamine and diethanolamine are particularly preferred.

[0044] Preferred examples of the nonionic surfactant include polyoxyalkylene alkyl(C₈₋₂₀) ethers, alkyl polyglycosides, polyoxyalkylene alkyl(C₈₋₂₀) phenyl ethers, polyoxyalkylene sorbitan fatty acid (C₈₋₂₂) esters, polyoxyalkylene glycol fatty acid (C₈₋₂₂) esters, and polyoxyethylene polyoxypropylene block copolymers. Of these, particularly preferred nonionic surfactants are polyoxyalkylene alkyl ethers [having an HLB number (as calculated by the Griffin method) of from 10.5 to 15.0, preferably from 11.0 to 14.5] obtained by adding 4 to 20 moles of an alkylene oxide such as ethylene oxide or propylene oxide to a C₁₀₋₁₈ alcohol.

[0045] The total amount of the surfactants in the detergent composition is preferably from 10 to 60 wt.%, more preferably 15 to 50 wt.%, still more preferably from 20 to 45 wt.% in view of detergency and solubility.

[0046] The amount of the anionic surfactant is preferably from 1 to 60 wt.%, more preferably from 1 to 50 wt.%, still more preferably from 3 to 40 wt.%, particularly in the powdery detergent composition.

[0047] The amount of the nonionic surfactant is preferably from 0.5 to 45 wt.%, more preferably from 1 to 35 wt.%, still more preferably from 3 to 25 wt.%, particularly in the powdery detergent composition.

[0048] The anionic surfactant and nonionic surfactant can be used singly, but a mixture thereof is preferred. In addition, an amphoteric surfactant or cationic surfactant may be used in combination according to the using purpose.

[0049] As a builder can be used such builder that has no detergency by itself or, if any, has no significant detergency, but can improve a detergent performance remarkably when incorporated in a detergent composition, particularly such builder that can improve the detergency of the surfactant which is a main component of the detergent composition. The builder must have, in addition, at least any one of multivalent metal cation scavenging action, stain dispersing action and alkali buffering action.

[0050] Examples of such a builder include water soluble inorganic compounds, water insoluble inorganic compounds and organic compounds.

[0051] Examples of the water soluble inorganic compounds include phosphates (such as tripolyphosphates, pyrophosphates, metaphosphates and trisodium phosphate), silicates, carbonates and sulfates. Of these, phosphates are preferred, because they have all the above-described three actions.

[0052] Examples of the water insoluble inorganic compounds include aluminosilicates (such as zeolite type A, zeolite type B, zeolite type X, and amorphous aluminosilicates), and crystalline silicates. Of these, zeolite type A having a particle size of 3 µm or less (more preferably, 1 µm or less) is preferred.

[0053] Examples of the organic compounds include carboxylates (such as aminocarboxylates, hydroxyaminocarboxylates, hydroxycarboxylates, cyclocarboxylates, maleic acid derivatives, and oxalates), and polymers of organic carboxylic acid (salt) (such as polymers and copolymers of acrylic acid, polymers and copolymers of polycarboxylic acid, polymers of glyoxylic acid, and polysaccharides and salts thereof). Of these, polymers of organic carboxylic acid

(salt) are preferred.

[0054] In the salts of the above-described builders, preferred counter ions are alkali metal salts and amines, with sodium and/or potassium, monoethanolamine and diethanolamine being particularly preferred.

[0055] Although the above-described builders may be used either singly or in combination, use of the water soluble inorganic compound is preferred, combined use of the water soluble inorganic compound and organic compound is more preferred, and combined use of the water soluble inorganic compound, organic compound and water insoluble inorganic compound is still more preferred.

[0056] The total content of the builders in the detergent composition is preferably from 20 to 80 wt.%, more preferably from 30 to 70 wt.%, still more preferably from 35 to 60 wt.% from the viewpoint of detergent performance.

[0057] The content of the water soluble inorganic compound contained as a builder, particularly in the powdery detergent composition, preferably ranges from 10 to 50 wt.%, more preferably from 15 to 45 wt.%, still more preferably from 20 to 40 wt.%.

[0058] The content of the water insoluble inorganic compound as a builder, particularly in the powdery detergent composition, preferably ranges from 5 to 50 wt.%, more preferably from 10 to 45 wt.%, still more preferably from 15 to 40 wt.%.

[0059] The content of the organic compound as a builder, particularly in the powdery detergent composition, preferably ranges from 0.1 to 20 wt.%, more preferably from 0.3 to 15 wt.%, still more preferably from 0.5 to 10 wt.%.

[0060] In the detergent composition of the present invention, additives such as bleaching agent (percarbonate, perborate, bleaching activator, etc.), anti-redepositioning agent (carboxymethyl cellulose, etc.), softening agents (dialkyl type quaternary ammonium salt, clay mineral, etc.), reducing agent (sulfite, etc.), fluorescent brighteners (biphenyl type, aminostilbene type, etc.), foam controlling agents (silicone, etc.) and perfumes can be incorporated.

[0061] In the detergent composition of the present invention, various enzymes in addition to the alkaline cellulase of the present invention may be incorporated. Examples of the enzyme include hydrolases, oxidases, reductases, transferases, lyases, isomerases, ligases and synthetases. Of these, cellulases other than those of the present invention, proteases, keratinases, esterases, cutinases, amylases, lipases, pullulanases, pectinases, mannanases, glucosidases, glucanases, cholesterol oxidases, peroxidases, and laccases are preferred. Among them, proteases, cellulases, amylases and lipases are especially preferred.

[0062] The detergent composition of the present invention can be prepared in a conventional manner by using the alkaline cellulase variant of the present invention obtained by the above-described method in combination with the above-described known detergent components. The form of the detergent can be selected according to the purpose of their use. It can be provided, for example, in the form of liquid, powder, granule, paste or solid.

[0063] The detergent composition of the present invention thus obtained can be used as a laundry powdery detergent, laundry liquid detergent, automatic dish washing detergent or cotton fiber modifying detergent.

[0064] An alternative embodiment of the present invention is directed to the production of an alkaline cellulase variant of the invention. Said method comprises the steps of:

(a) expression of a gene of the invention in a host under conditions which allow the expression at the encoded gene product, and

(b) recovery of the expressed alkaline protease.

[0065] Examples for corresponding hosts comprise prokaryotic organisms as well as eukaryotic cells and organisms. Preferably, the host is a microorganism. Appropriate conditions which allow the expression of the encoded gene product in host cultures, differ dependent from the used host. However, corresponding conditions are known to the person skilled in the art and described e.g. in several laboratory manuals such as Gerhard et al. "Methods for General and Molecular Bacteriology", American Society of Microbiology or Sambrook et al. "Molecular Cloning", Cold Spring Harbour Laboratory". Methods for the recovery of an expressed gene product from a culture are known to the person skilled in art from the field of biochemistry and also described in several laboratory manuals. One example of a corresponding method is described in detail in the appending examples.

[0066] A further alternative embodiment of the invention is directed to the use of an alkaline cellulase variant for the production of a detergent composition wherein the detergent composition is a laundry detergent or a cotton fiber modifying detergent. Preferably the laundry detergent is selected from the group of laundry powdery detergent, laundry liquid detergent and automatic dish washing detergent.

Examples

Example 1: Preparation of a variant of an alkaline cellulase gene

[0067] First, in order to obtain an alkaline cellulase variant permitting an increase in the production amount of the

alkaline cellulase, random mutagenesis in a region of the alkaline cellulase gene was conducted by the Error-Prone PCR method, whereby the library of the variants was constructed. From the variants thus obtained, a variant effective for improving the production amount of the alkaline cellulase was selected. After determination of the mutation sites by nucleotide sequencing, a multiple variant was constructed by random mutagenesis or site-specific mutagenesis at the mutation sites by using appropriate primers. As the template DNA, an alkaline cellulase gene derived from the *Bacillus* sp. strain KSM-S237 introduced in a plasmid, pHY300PLK, was employed.

[0068] As the mix primers for random mutagenesis were employed Ile16X (Primer 1, SEQ. ID NO:3, X means another amino acid), Phe39X (Primer 2, SEQ. ID NO:4), Ile76X (Primer 3, SEQ. ID NO:5), Met109X (Primer 4, SEQ. ID NO:6), Phe263X (Primer 5, SEQ. ID NO:7), Thr308X (Primer 6, SEQ. ID NO:8), Asn462X (Primer 7, SEQ. ID NO:9), Lys466X (Primer 8, SEQ. ID NO:10), Val468X (Primer 9, SEQ. ID NO:11), Ile552X (Primer 10, SEQ. ID NO:12), Ile564X (Primer 11, SEQ. ID NO:13) and Ser608X (primer 12, SEQ. ID NO:14).

[0069] The site-specific mutation introducing primers were employed, Leu10Gln (Primer 13, SEQ. ID NO:15), Ser22Pro (Primer 14, SEQ. ID NO:16), Asn33His (Primer 15, SEQ. ID NO:17), Phe39Ala (Primer 16, SEQ. ID NO:18), Met109Ile (Primer 17, SEQ. ID NO:19), Gln242Ser (Primer 18, SEQ. ID NO:20), Gln242Ala (Primer 19, SEQ. ID NO:21), Gln242Phe (Primer 20, SEQ. ID NO:22), Gln242Val (Primer 21, SEQ. ID NO:23), Gln242Asp (Primer 22, SEQ. ID NO:24) or Gln242Glu (Primer 22, SEQ. ID NO:24), Gln242Gly (Primer 23, SEQ. ID NO:25), Gln242Ile (Primer 20, SEQ. ID NO:22), Gln242Thr (Primer 24, SEQ. ID NO:26), Gln242Leu (Primer 25, SEQ. ID NO:27), Gln242Met (Primer 26, SEQ. ID NO:28), Gln242Tyr (Primer 27, SEQ. ID NO:29), Phe263Ile (Primer 28, SEQ. ID NO:30), Thr308Ala (Primer 29, SEQ. ID NO:31) and Ile552Met (Primer 30, SEQ. ID NO:32).

[0070] Specifically, after mixing 0.5 μ L (10 ng) of the template DNA plasmid, 20 μ L (1 μ M) of the mutation introducing primer, 20 μ L (1 μ M) of the antisense primer, 10 μ L of a $\times 10$ PCR buffer solution, 8 μ L of a 10 mM deoxynucleotide triphosphate (dNTP) mixture, 0.5 μ L (2.5 units) of "Pyrobest DNA polymerase" (product of Takara) and 39.5 μ L of deionized water, PCR was done by "Gene Amp PCR system 9700" (product of Amesham-Pharmacia). The reaction condition was 94°C for 2 minutes, followed by 30 cycles of 94°C for 1 minute, 56°C for 1 minute, and 72°C for 30 seconds, and finally 72°C for 1 minute. After purification of the resulting PCR product by "GFX PCR DNA and Gel Band Purification Kit" (Amesham-Pharmacia), 5.5 μ L of a $\times 10$ phosphorylation buffer and 1 μ L (10 units) of polynucleotide kinase were added to the solution, and it was incubated at 37°C for 1 hour (50 μ L). After mixing 25 μ L of the phosphorylated PCR product with 2 μ L (20 ng) of the template plasmid, 10 μ L of a $\times 10$ PCR buffer, 8 μ L of a 10 mM dNTP mixture, 1 μ L (5 units) of "Pyrobest DNA polymerase" and 54 μ L of deionized water, PCR was conducted. The reaction condition was 94°C for 2 minutes, followed by 30 cycles of 94°C for 1 minute, 60°C for 1 minute and 72°C for 6 minutes, and finally 72°C for 12 minutes.

[0071] The resulting PCR product was purified (43.5 μ L). Then, 5.5 μ L of a $\times 10$ phosphorylation buffer and 1 μ L (10 units) of polynucleotide kinase were added, and phosphorylation was conducted at 37°C for 1 hour. The mixture was subjected to ethanol precipitation. The DNA solution (10 μ L) thus collected was subjected to ligation at 16°C for 18 hours by using a ligation kit ver. 2 (product of Takara), followed by ethanol precipitation again, whereby the DNA mixture was collected.

Example 2

[0072] By using 5 μ L of the DNA mixture obtained in Example 1, the DNA was introduced into the *Bacillus subtilis* strain ISW1214 according to Chang and Cohen (Mol. Gen. Gent., 168, 111-115, 1979) whereby the corresponding transformant was obtained. Specifically, after shaking *B. subtilis* strain ISW1214 in a 50 mL of LB medium at 37°C for about 2 hours (absorbance at 600 nm: 0.4), the cells were collected by centrifugal separation (at 7000 rpm for 15 minutes) at room temperature and suspended in a 5 mL of SMMP [0.5M sucrose, 20 mM disodium maleate, 20 mM magnesium chloride 6 hydrate, 35% (w/v) "Antibiotic Medium 3" (Difco)]. To the resulting suspension was added 500 μ L of a lysozyme solution (30 mg/mL) dissolved in an SMMP solution, and the resulting solution was incubated at 37°C for 1 hour. Then, protoplasts were collected by centrifugal separation (at 2800 rpm, for 15 minutes) at room temperature and suspended in 5 mL of SMMP to give a protoplast solution. To 0.5 mL of the protoplast solution were added 10 μ L of a plasmid solution and 1.5 mL of a 40% (w/v) polyethylene glycol (PEG8,000, Sigma). The resulting mixture was stirred slowly. After allowing to stand at room temperature for 2 minutes, the mixture was mixed with 5 mL of an SMMP solution. By centrifugal separation (at 2800 rpm for 15 minutes) at room temperature, protoplasts were collected and suspended again in 1 mL of an SMMP solution. The protoplast suspension was shaken at 37°C for 90 minutes (at 120 rpm), then applied onto a DM3 regeneration agar medium [0.8% (w/v) agar (product of Wako Pure Chemicals), 0.5% disodium succinate 6 hydrate, 0.5% "Casamino Acids Technical" (product of Difco), 0.5% yeast extract, 0.35% monopotassium phosphate, 0.15% dipotassium phosphate, 0.5% glucose, 0.4% magnesium chloride 6 hydrate, 0.01% bovine serum albumin (product of Sigma), 0.5% carboxymethyl cellulose, 0.005% trypan blue (product of Merck) and an amino acid mixture (leucine and methionine, each 10 μ g/mL)] containing tetracycline (15 μ g/mL, Sigma), and incubated at 30°C for 72 hours to give a transformant. The transformant having a halo formed on the DM3 regeneration agar

plate was subjected to shake cultivation at 30°C for 15 hours in a polypeptone medium containing tetracycline (15 µg/mL). After collection of the cells, plasmids were collected and purified by "Micro Prep Plasmid Purification kit" (product of Amesham-Pharmacia).

Example 3: Determination of nucleotide sequence

[0073] The nucleotide sequence of the cellulase gene inserted in the plasmids obtained in Example 2 was confirmed using "377DNA Sequencer" (product of Applied Biosystems).

Example 4: Evaluation of the production of cellulase variant

[0074] The host bacteria *B. Subtilis* strain ISW1214 was cultured in a medium (PSM medium) containing 3% (W/v) "Polypepton S" (product of Nihon Pharmaceutical), 0.5% fish meat extract (Wako Pure Chemicals), 0.05% yeast extract, 0.1% monopotassium phosphate, 0.02% magnesium sulfate 7 hydrate, tetracycline (15 µg/mL) and 5% maltose at 37°C for 72 hours.

[0075] The activity in the culture supernatant of each cellulase variant was assayed. Assuming that the production amount of the recombinant wild cellulase per volume of culture is 100%, the production amount of the alkaline cellulase variant Leu10Gln was 120%, Ile16Asn 139%, Ser22Pro 140%, Asn33His 105%, Phe39Ala 113%, Ile76His 112%, Met109Leu 112%, Gln242Ser 125%, Phe263Ile 137%, Thr308Ala 102%, Asn462Thr 116%, Lys466Leu 110%, Val468Ala 122%, Ile552Met 132%, Ile564Val 113%, and Ser608Ile 110%. The production amount of a double variant Leu10Gln + Ser22Pro was 117%, Asn33His + Phe39Ala 118%, Ser22Pro + Gln242Ala 207%, Ser22Pro + Gln242Phe 196%, Ser22Pro + Gln242Val 187%, Ser22Pro + Gln242Ser 175%, Ser22Pro + Gln242Ile 174%, Ser22Pro + Gln242Gly 160%, Ser22Pro + Gln242Glu 158%, Ser22Pro + Gln242Asp 145%, Ser22Pro + Gln242Thr 134%, Ser22Pro + Gln242Leu 128%, Ser22Pro + Gln242Met 125%, and Ser22Pro + Gln242Tyr was 115%. The production amount of a triple variant Leu10Gln + Ser22Pro + Gln242Ser was 162% and that of Ile16Asn + Ser22Pro + Gln242Ser was 107%. The production amount of a quadruple variant Leu10Gln + Ser22Pro + Ile76His + Lys466Leu was 113%, while that of a quintuple variant Leu10Gln + Ile16Asn + Ile76His + Gln242Ser + Lys466Leu was 163%, thus showing an improvement in the production amount.

[0076] Analysis results have revealed that an improvement in the production amount of most of the variants owes to an increase in the secretion amount of protein. The variants substituted at position 242 were recognized to exhibit an improvement in specific activity against carboxymethyl cellulose (CMC) serving as a substrate. Specifically, assuming that the specific activity of a wild type enzyme is 100%, the specific activity of Gln242Ala was 147%, Gln242Val 150%, Gln242Ser 125%, Gln242Phe 128%, Gln242Asp 107%, Gln242Glu 106% and Gln242Gly 105%, thus showing an improvement in a relative specific activity. Moreover, the specific activity in a 0.1M phosphate buffer (pH 8) also showed an improvement. For example, the specific activity of Gln242Ala was 150%, Gln242Ser 110%, Gln242Val 123%, Gln242Phe 110%, Gln242Asp 108% and Gln242Glu 114%, thus showing an improvement in the relative specific activity. The protein content was determined by "Protein Assay Kit" (product of Bio-Rad) using bovine serum albumin as the standard protein.

<Cellulase Assay (3,5-dinitrosalicylic acid (DNS) method)>

[0077] To a reaction mixture composed of 0.2 mL of a 0.5M glycine-sodium hydroxide buffer (pH 9.0), 0.4 mL of 2.5% (w/v) carboxymethyl cellulose (A01MC: Nippon Paper Industries), and 0.3 mL of deionized water was added 0.1 mL of a properly diluted enzyme solution. After the resulting mixture was incubated at 40°C for 20 minutes, 1 mL of a dinitrosalicylic acid reagent (0.5% dinitrosalicylic acid, 30% Rochelle Salt, 1.6% sodium hydroxide) was added to develop the color of a reducing sugar in a boiling water for 5 minutes. After quenching in ice water, 4 mL of deionized water was added and absorbance at 535 nm was measured to determine the production amount of the reducing sugar. A blank test was conducted in a similar manner excepting that the enzyme solution was added just before incubation in a boiling water bath. One unit (1U) of an enzyme activity was defined as an amount of protein which produces a reducing sugar in an amount equivalent to 1 µmol of glucose in 1 minute under the above-described reaction conditions.

Example 5

[0078] A powdery detergent having the below-described composition made of A-1 particles, C-1 particles and perfume as described in Japanese Patent Laid-Open No. 2002-265999 was prepared and after addition of the alkaline cellulase variant of the present invention in an amount of 1350000 U/kg, the detergency of the enzyme-containing detergent was evaluated.

Table 2

(Component)	(wt.%)
A-1 particles	99
C-1 particles	0.5
Perfume	0.5
Total	100

<Detergency Measuring Method>

(Artificially stained cloth)

[0079] An artificially stained cloth for detergency evaluation as described in Japanese Patent Laid-Open No. 2002-265999 was employed.

(Washing conditions, Washing method and Evaluation method)

[0080] A detergent was dissolved in hard water (CaCl_2 : 55.42 mg/L, $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$: 43.51 mg/L) at 30°C to prepare 1L of a 0.00667 wt.% aqueous solution of the detergent. Five pieces of artificially stained cloth were immersed in the aqueous detergent solution at 30°C for 1 hour, followed by stirring and washing in a Terg-O-Tometer at 100 rpm, 30°C for 10 minutes. After rinsing each cloth with running water, it was iron-pressed and provided for the measurement of reflectance. The reflectance at 550 nm of the cloth before staining, and those of the artificially stained cloth before and after washing were measured by an automatic colorimeter (product of Shimadzu Corporation). In accordance with the below-described equation, a washing ratio (%) was determined and detergency was evaluated based on the average of the washing ratios of the five cloths.

Washing ratio(%) =

$$\frac{\text{Reflectance of stained cloth after washing} - \text{Reflectance of stained cloth before washing}}{\text{Reflectance of cloth before staining} - \text{Reflectance of stained cloth before washing}} \times 100$$

(Evaluation results)

[0081] As a result, the detergent containing the alkaline cellulase variant of the present invention exhibited a washing ratio of 71%, which correspond to 7% increase compared with the washing ratio 64% of a cellulase free detergent.

Industrial Applicability

[0082] When the alkaline cellulase variant of the present invention is employed, a culture broth having a higher activity compared with the conventional one is available, which makes it possible to supply a large amount of an alkaline cellulase for industrial uses including detergent. When a predetermined amount of an enzyme is produced, the number of times of cultivation can be reduced, whereby decreasing the energy necessary for cultivation, amounts of medium components, a carbon dioxide amount formed during cultivation and an amount of water discharged.

SEQUENCE LISTING

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20 25 30

Asn Thr Arg Glu Asp Asn Phe Lys His Leu Leu Gly Asn Asp Asn Val

35 40 45

Lys Arg Pro Ser Glu Ala Gly Ala Leu Gln Leu Gln Glu Val Asp Gly

50 55 60

Gln Met Thr Leu Val Asp Gln His Gly Glu Lys Ile Gln Leu Arg Gly

65 70 75 80

Met Ser Thr His Gly Leu Gln Trp Phe Pro Glu Ile Leu Asn Asp Asn

85 90 95

Ala Tyr Lys Ala Leu Ser Asn Asp Trp Asp Ser Asn Met Ile Arg Leu

100 105 110

Ala Met Tyr Val Gly Glu Asn Gly Tyr Ala Thr Asn Pro Glu Leu Ile

115 120 125

Lys Gln Arg Val Ile Asp Gly Ile Glu Leu Ala Ile Glu Asn Asp Met

130 135 140

Tyr Val Ile Val Asp Trp His Val His Ala Pro Gly Asp Pro Arg Asp

145 150 155 160

Pro Val Tyr Ala Gly Ala Lys Asp Phe Phe Arg Glu Ile Ala Ala Leu

165 170 175

Tyr Pro Asn Asn Pro His Ile Ile Tyr Glu Leu Ala Asn Glu Pro Ser

180 185 190

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EP 1 350 843 A2

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	325	330	335	
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	Ala Asn Trp Ser Leu Thr Asn Lys Asn Glu Val Ser Gly Ala Phe Thr			
	340	345	350	
55	cca ttc gag tta ggt aag tct aac gca acc aat ctt gac cca ggt cca			1104
	Pro Phe Glu Leu Gly Lys Ser Asn Ala Thr Asn Leu Asp Pro Gly Pro			
	355	360	365	

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 Asp His Val Trp Ala Pro Glu Glu Leu Ser Leu Ser Gly Glu Tyr Val
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10

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35

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40

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45

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EP 1 350 843 A2

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 15 770 775 780

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Claims

1. An alkaline cellulase variant obtained by substituting the amino acid residue of a cellulase having an amino acid sequence exhibiting at least 90 % homology with the amino acid sequence represented by SEQ ID NO: 1 at (a) position 10, (b) position 16, (c) position 22, (d) position 33, (e) position 39, (f) position 76, (g) position 109, (h) position 242, (i) position 263, (j) position 308, (k) position 462, (l) position 466, (m) position 468, (n) position 552, (o) position 564, or (p) position 608 in SEQ ID NO: 1, or at a position corresponding thereto has been selected from the following amino acid residue.
2. An alkaline cellulase variant of a cellulase having an amino acid sequence exhibiting at least 90 % homology with the amino acid sequence represented by SEQ ID NO: 1 and substitutions of the amino acid residues with another amino acid residue at (a) position 10, (b) position 16, (c) position 22, (d) position 33, (e) position 39, (f) position 76, (g) position 109, (h) position 242, (i) position 263, (j) position 308, (k) position 462, (l) position 466, (m) position 468, (n) position 552, (o) position 564, or (p) position 608 in SEQ ID NO: 1, or at a position corresponding thereto has been selected from the following amino acid residue.
3. An alkaline cellulase variant of claim 1 or 2, wherein the another amino acid residue is selected from the following amino acid residues:
 - (a) at position 10: glutamine, alanine, proline or methionine
 - (b) at position 16: asparagine or arginine
 - (c) at position 22: proline
 - (d) at position 33: histidine
 - (e) at position 39: alanine, threonine or tyrosine
 - (f) at position 76: histidine, methionine, valine, threonine or alanine
 - (g) at position 109: isoleucine, leucine, serine or valine
 - (h) at position 242: alanine, phenylalanine, valine, serine, aspartic acid, glutamic acid, leucine, isoleucine, tyrosine, threonine, methionine or glycine
 - (i) at position 263: isoleucine, leucine, proline or valine
 - (j) at position 308: alanine, serine, glycine or valine
 - (k) at position 462: threonine, leucine, phenylalanine or arginine
 - (l) at position 466: leucine, alanine or serine
 - (m) at position 468: alanine, aspartic acid, glycine or lysine
 - (n) at position 552: methionine
 - (o) at position 564: valine, threonine, or leucine, and

(p) at position 608: isoleucine or arginine.

4. A gene encoding an alkaline protease as claimed in claim 1, 2 or 3.

5. A recombinant vector comprising a gene as claimed in claim 4.

6. A transformant comprising a vector as claimed in claim 5.

7. A transformant of claim 6, wherein a microorganism is used as a host.

8. A detergent composition comprising an alkaline cellulase variant of claim 1, 2 or 3.

9. A method for the production of an alkaline cellulase variant according to claim 1, 2 or 3 said method comprising the steps of:

(a) expression of a gene according to claim 4 in a host under conditions which allow the expression at the encoded gene product, and

(b) recovery of the expressed alkaline protease.

10. The use of alkaline cellulase variant for the production of a detergent composition wherein the detergent composition is a laundry detergent or a cotton fiber modifying detergent.

```

Egl-237 1:MMLRKKTQOLISSILILVLLSLFPALAAEGNTREDNFKHLIGNDNVKRPSEAGALQLOEVDGQMTLVDQHEKIQLRGMSHGLQWFP 90
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Egl-64 1:MMLRKKTQOLISSILILVLLSLFPALAAEGNTREDNFKHLIGNDNVKRPSEAGALQLOEVDGQMTLVDQHEKIQLRGMSHGLQWFP 90
Egl-N131b 1:MMLRKKTQOLG-----PAQA--EGNTREDNFKHLIGNDNVKRPSEAGALQLOEVDGQMTLVDQHEKIQLRGMSHGLQWFP 76
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Figure 1

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Eg1-N131b 347: ATSLDPGDPQVWVPEELSLSGEYVRARIKGVNYEPIDRTKYTKVLMDFNDGKQGFVNSDSPNKELIADVNNNTLKVSGLDVSNVSD 436
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Figure 1

28

Figure 1

(19)



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(11)

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(12)

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(54) **Alkaline cellulase variants**

(57) Provided in the present invention are an alkaline cellulase variant obtained by using a cellulase having an amino acid sequence exhibiting at least 90% homology with the amino acid sequence represented by SEQ. ID NO:1 and substituting the amino acid residue of the cellulase at (a) position 10, (b) position 16, (c) position 22, (d) position 33, (e) position 39, (f) position 76, (g) position 109, (h) position 242, (i) position 263, (j) position 308, (k) position 462, (l) position 466, (m) position 468, (n) position 552, (o) position 564, or (p) position

608 in SEQ. ID NO:1, or at a position corresponding thereto with another amino acid residue; a gene encoding the variant; a vector containing the gene; a transformant containing the vector; and a detergent composition containing the alkaline cellulase variant.

The present invention makes it possible to provide an alkaline cellulase which acts favorably in an alkaline range, and can be mass produced easily because it has a high secretion capacity or enhanced specific activity.

EP 1 350 843 A3



European Patent
Office

EUROPEAN SEARCH REPORT

Application Number
EP 03 00 6842

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.7)
X	DATABASE GENESEQ [Online] Derwent; 8 February 2002 (2002-02-08) Database accession no. AAG80267 XP002251476	1,2,4-10	C12N9/42 C11D3/386
Y	* the whole document * & JP 2001 231569 A (KAO CORP.) 28 August 2001 (2001-08-28) ---	3	
X	DATABASE GENESEQ [Online] Derwent; 2 February 1993 (1993-02-02) Database accession no. AAR26021 XP002251477	1,2,4-10	
Y	* abstract * & JP 04 190793 A (KAO CORP.) 9 July 1992 (1992-07-09) ---	3	
Y	WO 98 12307 A (CHRISTIANSEN LARS ;DAMGAARD BO (DK); NOVONORDISK AS (DK); ANDERSEN) 26 March 1998 (1998-03-26) * the whole document * -----	3	
			TECHNICAL FIELDS SEARCHED (Int.Cl.7)
			C12N C11D
The present search report has been drawn up for all claims			
Place of search MUNICH		Date of completion of the search 18 August 2003	Examiner Wimmer, G
<p>CATEGORY OF CITED DOCUMENTS</p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons</p> <p>& : member of the same patent family, corresponding document</p>			

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